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CheKine™ Micro Formaldehyde Dehydrogenase (FDH) Activity Assay Kit

Cat #: KTB3033 Size: 48 T/96 T

[-]	Micro Formaldehyde Dehydrogenase (FDH) Activity Assay Kit				
REF	Cat #: KTB3033	LOT	Lot #: Refer to product label		
	Applicable samples: Animal and Plant Tissues, Cells, Serum, Plasma or other Liquids				
Ĵ.	Storage: Stored at -20°C for 6 months, protected from light				

Assay Principle

Formaldehyde is a non-specific reactive compound with proteins, nucleic acids and lipids, and is highly toxic to all organisms. As one of the family members of Zinc-containing medium chain alcohol dehydrogenase (ADH), formaldehyde dehydrogenase is widely present in prokaryotes and eukaryotes. This enzyme can use NAD⁺ as a coenzyme to oxidize toxic formaldehyde and is a key enzyme in the formaldehyde oxidation pathway. FDH catalyzes formaldehyde and NAD⁺ to produce NADH, and the absorbance value at 340 nm increases. The activity of FDH can be reflected by the change of absorbance value of NADH at 340 nm.

Materials Supplied and Storage Conditions

Kit components	Size		Storage conditions
Tat components	48 T	96 T	
Extraction Buffer	50 mL	100 mL	4℃
Reagent	7.5 mL	15 mL	4℃
Reagent II	1	1	-20°C
ReagentIII	1	1	4°C, protected from light
Reagent IV	0.75 mL	1.5 mL	4°C, protected from light

Materials Required but Not Supplied

- · Microplate reader or ultraviolet spectrophotometer capable of measuring absorbance at 340 nm
- 96-well UV plate or microquartz cuvette, precision pipettes, disposable pipette tips
- · Water bath, ice maker, centrifuge
- Deionized water
- Homogenizer (for tissue samples)



Reagent Preparation

Extraction Buffer: Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C.

Reagent I: Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C.

Reagent II: Prepare before use, 48 T add 3 mL deionized water, 96 T add 6 mL deionized water to dissolve it for use. The unused Reagent II can be stored at -20°C for one month after packaging.

Reagent III: Prepare before use, 48 T add 0.75 mL deionized water, 96 T add 1.5 mL deionized water to dissolve it for use. The unused Reagent III can be stored at 4°C for one month, protected from light.

Reagent IV: Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C, protected from light.

Note: Reagent IV is toxic, please wear protective measures such as mask and gloves during the experiment.

Sample Preparation

Note: It is recommended to use fresh samples. If the experiment is not conducted immediately, the samples can be stored at -80°C for 1 month. The temperature and time of thawing should be controlled during the determination. When thawing at room temperature, the sample should be thawed within 4 h.

- 1. Tissues: Weigh 0.1 g tissue, add 1 mL Extraction Buffer, homogenize on ice. Centrifuge at 10,000 g for 20 min at 4°C. Take the supernatant and place it on the ice for testing.
- 2. Cells: Collect 5×10⁶ cells into the centrifuge tube, wash cells with cold PBS, discard the supernatant after centrifugation, add 1 mL Extraction Buffer, ultrasonically disrupt cells 3 min (power 300 W, ultrasonic 3 s, interval 7 s, total time for 3 min). Then centrifuge at 10,000 g for 20 min at 4°C. Take the supernatant and place it on the ice for testing.
- 3. Serum, Plasma and other Liquid Samples: Direct detection. If the solution has turbidity, centrifuge and take the supernatant for measurement.

Note: If the protein concentration of the sample is need to determined, it is recommended to use Abbkine catalog number: KTD3001 Protein Quantification Kit (BCA Assay) to measure the protein concentration of the sample.

Assay Procedure

- 1. Preheat the microplate reader or ultraviolet spectrophotometer for more than 30 min, and adjust the wavelength to 340 nm, ultraviolet spectrophotometer was returned to zero with deionized water.
- 2. Preheat the prepared Reagent || at 37°C for 5 min.
- 3. Sample measurement. (The following operations are operated in the 96-well UV plate or microquartz cuvette)

Regent	Test Well (µL)	
Sample	20	
Reagent	110	
Reagent	50	
ReagentIII	10	
ReagentIV	10	

Mix well, record the absorbance values of 0 min and 30 min at 340 nm, mark as A_1 and A_2 , and calculate $\Delta A = A_2 - A_1$.

Note: In order to guarantee the accuracy of experimental results, need to do a pre-experiment with 2-3 samples. If the number of samples is large, the reagents can be proportionally mixed into a working regent for use. If the ΔA is less than 0.01, the sample size can be appropriately increased or the reaction time can be appropriately extended (For example, the reaction time is 60 min). If the ΔA is greater than 1.0, the sample can be appropriately diluted with Extraction Buffer, the calculated result multiplied by the dilution factor.



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Data Analysis

Note: We provide you with calculation formulae, including the derivation process and final formula. The two are exactly equal. It is suggested that the concise calculation formula in bold is final formula.

A. 96-well UV plates calculation formula

(1) Calculated by protein concentration:

Unit definition: One enzyme activity unit defines as 1 nmol NADH produced by 1 mg tissue proteins per min.

FDH (U/mg prot)= $[\Delta A \times V_{Total} \div (\epsilon \times d) \times 10^9] \div (V_{Sample} \times Cpr) \div T = 128.6 \times \Delta A \div Cpr$

(2) Calculation according to the weight of the sample:

Unit definition: One enzyme activity unit defines as 1 nmol NADH produced by 1 q tissue per min

FDH (U/g fresh weight)= $[\Delta A \times V_{Total} \div (\epsilon \times d) \times 10^9] \div (V_{Sample} \times W \div V_{Sample Total}) \div T$ =128.6× $\Delta A \div W$

(3) Calculation according to cell number

Unit definition: One enzyme activity unit defines as 1 nmol NADH produced by 10⁴ cells per min

FDH (U/10⁴ cell)= $\Delta A \div (\epsilon \times d) \times V_{Tota} \div (V_{Sample} \times 500 \div V_{Sample} \times 100 \div V_$

(4) Calculation according to the volume of liquid

Unit definition: One enzyme activity unit defines as 1 nmol NADH produced by 1 mL liquid per min

FDH (U/mL)= $[\Delta A \times V_{Total} \div (\epsilon \times d) \times 10^{9}] \div V_{Sample} \div T = 128.6 \times \Delta A$

Where: ϵ : NADH molar extinction coefficient, 6.22×10³ L/mol/cm; d: 96-well UV plate diameter, 0.5 cm; V_{Total}: the total volume of the reaction system, 0.2 mL=2×10⁻⁴ L; V_{Sample}: the volume of the sample in the reaction system, 0.02 mL; V_{Sample Total}: The volume of Extraction Buffer added, 1 mL; Cpr: protein concentration (mg/mL); W: sample weight, g; T: reaction time, 30 min; 500: Total number of cells, 5×10^6 .

B. Microquartz cuvette calculation formula

The optical diameter d:0.5 cm in the above calculation formula can be adjusted to d:1 cm for calculation.

Typical Data

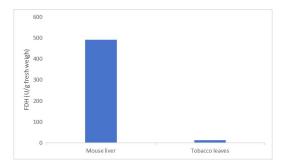


Figure 1. FDH activity in Mouse liver and Tobacco leaves was detected with this kit.

Recommended Products

Catalog No.	Product Name		
KTB1015	CheKine™ Micro α-glucosidase(α-GC) Activity Assay Kit		
KTB1121	CheKine™ Micro Pyruvate Acid (PA) Assay Kit		

Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes.



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